



Calendula Officinalis enhance the wound healing potential in diabetic rats

Saravanan VS¹, Kalpana Krishnaraju¹, Manimekalai Pichaivel², Krishnaraju Venkatesan^{3*}, Premalatha Paulsamy⁴, Divya Kuppan⁵

¹Erode College of Pharmacy, Veppampalayam, Erode, India

²Professor & Head Department of Pharmacology, SVCP, Tiruchengode, Tamilnadu, India

^{3*}Department of Pharmacology, College of Pharmacy, King Khalid University, Abha, KSA

⁴King Khalid University, Khamis Mushayit, Asir Province, Saudi Arabia.

⁵JKK Nataraja College of arts and science, Kumarapalayam, Namakkal District, India

Received: 09-06-2021 / Revised Accepted: 27-07-2021 / Published: 27-07-2021

ABSTRACT

The goal of this study was to see how well *Calendula.officinalis* (*C.officinalis*) could treat dead space wounds in rats in vivo. On each axilla of diabetic rats, dead space incisions were created. For eight days, the rats were randomly assigned to one of three treatment groups (Group I: Normal saline; Group II: Diabetic control; Group III: Tannin). Animals were euthanized on day 10, and cotton pellets and granuloma tissues were carefully collected and processed for further estimates. Tissue breaking strength, dry and wet weight, and biochemical markers including hydroxyproline, hesosamine, and tissue protein were used to assess healing capacity. The extract was taken orally and had a favourable effect on the wound. The *C.officinalis* extract shows considerable wound healing activity in diabetic wounds, based on the preceding findings.

Key words: *C.officinalis* , Wound healing; Diabetic; Dead space wound; Granulation tissue; Streptozotocin.

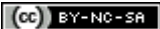
INTRODUCTION

Wounds are clinical entities that occur often in everyday life. A wound is a break in the continuity of live tissue caused by an injury. Wounds are inconvenient, and they are more prone to infection and other problems. Some illnesses, such as immune compromised states, ischaemia, and malnutrition, ageing, local infection, and local tissue injury, cause wound healing to be delayed.

Wound healing is a complex process in which the skin heals itself following an injury. Inflammatory, proliferative, and remodelling phases are the three stages of wound healing. Increased blood flow, increased capillary permeability, and increased leucocyte migration in the affected area characterise the inflammatory phase. Granulation, contraction, and epithelisation are all characteristics of the proliferative phase.

Address for Correspondence: Dr. Krishnaraju Venkatesan, Department of Pharmacology, College of Pharmacy, King Khalid University, Abha, KSA; Email: kvenkatesan@kku.edu.sa

How to Cite this Article: Saravanan VS, Kalpana Krishnaraju, Manimekalai Pichaivel, Krishnaraju Venkatesan, Premalatha Paulsamy, Divya Kuppan. *Calendula Officinalis* enhance the wound healing potential in diabetic rats. World J Pharm Sci 2021; 9(8): 147-150.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License, which allows adapt, share and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms. 

The strength and appearance of the healed region are determined during the remodelling phase.¹ *Calendula.officinalis* (*C.officinalis*) often known as pot marigold, is a common Asteraceae plant that is native to southern Europe. Phyto-chemicals found in the plant species include carbohydrates, phenolic compounds, lipids, steroids, tocopherols, terpenoids, quinones, and carotenoids²⁻⁵ all of which have various health effects⁶⁻⁸ and⁹ Triterpendiol esters, saponins, and flavonoids such as rutin and hyperoside are among the plant's most active components. This plant is utilised in the form of infusions, tinctures, liquid extracts, lotions, and ointments for medical purposes. This factory also produces skin care items that are sold all over the world.

The goal of this study was to see how effective aqueous extracts of *C.officinalis* were at healing wounds on excision-wounded rats. Wound healing is frequently hindered in people with diabetes mellitus (DM), resulting in non-healing, delayed healing, or persistent skin ulcers.¹⁰ In diabetes, delayed wound healing can be caused by an imbalance in the inflammatory response, changed cytokine production, altered collagen synthesis, insufficient angiogenesis, extracellular matrix differentiation, lower tensile strength, or diminished growth factors.^{11,12} Incision and excision wound models were used to investigate the effects of tannin on wound healing activities. The tannin impact on a diabetic wound model, on the other hand, is unknown. As a result, the goal of this study is to see how *C.officinalis* affects wound in streptozotocin (STZ) induced diabetes in rats.

MATERIALS AND METHODS

Preparation of extract: *C.officinalis* leaves (100g) were coarsely pulverized. In a Soxhlet extractor, the powdered materials were loaded and defatted using petroleum ether (40-60°C). The marc was dried and extracted three times with ethanol (50 percent v/v) in the same extractor. Finally, using a rotary evaporator under vacuum, the extracts were condensed to a semi solid mass. The solvent was removed from the dried extract by placing it in a desiccator.

Animals: Healthy Wistar rats of either sex (150–200 g) were utilised in this study, and no prior pharmacological therapy was given to them. The animals were fed a commercial pellet diet and given unlimited water. The animals were given a 10 day acclimatisation period before starting the experiment. The therapy was carried out with the approval of the animal ethics committee of King Khalid University and in compliance with the National Institute of Health's standards for the care and use of laboratory animals in the United States

(NIH Publication No. 85-23, revised 1996). For the dead space wound model, animals of either sex were divided into three groups, each with six animals: Group I- Normal control; group II-diabetic control; and group III received *C.officinalis* (100 mg/kg/day). The extracts were administered orally to the individual animal groups once a day.

Wound healing activity:

Dead Space wound model: Rathi et al. described a technique for creating dead space wounds.¹³ Eighteen rats were divided into three groups of six individuals each. Subcutaneous dead space wounds were produced in the area of the axilla by creating a pouch through a tiny nip in the skin under general anaesthesia (achieved with 10 mg/kg body weight of xylazine hydrochloride and 50 mg/kg ketamine hydrochloride). The development of granulomas was induced by implanting sterile cotton pellets (30 mg) in each axilla. Sutures were used to close the wounds, which were then cleaned with an alcoholic swab. After being grouped together, the animals were placed individually in a metal cage to prevent them from biting each other's wounds.

For 8 days, the treatment groups were given extract or normal saline (1 ml/kg). Rats were euthanized on day 10 and the cotton pellets and granuloma tissues were carefully removed, dried in a 60°C oven, weighed, and compared to the control. The hydroxyproline, hexosamine concentration, and hexuronic acid were determined using the neutralised acid hydrolyzate of the dry tissue. For the measurement of lysyl oxidase and tissue protein, a piece of the moist granulation tissue was utilised.

Induction of diabetes: The overnight starved rats were given a newly produced solution of streptozotocin (STZ) (Sigma, St. Louis, MO, USA) dissolved in citrate buffer pH 4.5 at a dosage of 65 mg/kg intraperitoneally (i.p.) 15 minutes after receiving 110 mg/kg body weight nicotinamide (HiMedia labs Pvt. Ltd.). After 6 hours of STZ treatment, the rats were given a 10% glucose solution for additional 24 hours to prevent hypoglycemia caused by large pancreatic insulin secretion. Blood was collected from the tail veins of the rats 72 hours after the STZ injection, and those with a fasting blood glucose level of more than 200 mg/dl were classified as diabetic and used in this investigation.¹⁴

Statistical analysis: The data is presented as a mean with a standard deviation (SEM). The differences between means were investigated using one way analysis of variance (ANOVA), with p values less than 0.05 being deemed significant. The data was analysed using one way analysis of variance (ANOVA) with a post hoc Scheffe's test in

Graph Pad, and the mean and standard deviation were calculated.

RESULTS

Animals given the *C.officinalis* extract showed a substantial increase in wound-healing activity when compared to those given sham treatments. The effects of *C.officinalis*, given orally at a dosage of

100 mg kg-1 day-1 for 8 days, on wound healing activity in rats with dead space wounds are shown in Table 1. When compared to diabetic and control rats, the *C.officinalis* treatment group's granulation tissue breaking strength and wet and dry granulation tissue weight were considerably higher (table-1).

Table-1: Physical and biochemical analysis of granulation tissue in streptozotocin induced diabetic rats

Groups	Blood glucose (mg/dl)	Wet weight (mg/100g rat)	tissue (mg/100g rat)	Dry weight (mg/100g rat)	tissue (mg/100g rat)	Tissue breaking strength (g)
Wounded Control	82.1 ± 6.0	245.5 ± 14.09		30.38 ± 5.20		275.19±15.17
Diabetic Control	276.38 ± 14.1 ^a	169.5 ± 10.32 ^a		26.5 ± 4.40 ^a		166.41±1.30 ^a
<i>C.officinalis</i>	281.18 ± 12.1 ^a	279.5 ± 12.19 ^a		30.5 ± 4.20 ^a		313.19±13.37 ^a

Values are mean ± SD of 6 replications. P values: ^a:<0.01vs control.

In diabetic rats produced by streptozotocin, the concentration of hydroxyproline in granulation tissue was substantially lower. The experimental group had considerably lower levels of glycosaminoglycan contents such as hexuronic acid and hexosamine. When diabetic rats were

compared to control rats, tissue protein content was extremely low. In the experimental group, the amount of lysyl oxidase was substantially lower. In comparison to diabetic and control rats (group II), all of the following metrics significantly increased in the *C.officinalis* treatment group (table 2).

Table 2: Biochemical analysis of granulation tissue in streptozotocin induced diabetic rats

Groups	Hydroxyproline (mg/g tissue)	Hexosamines (mg/g tissue)	Hexuronic acid (mg/g tissue)	Tissue protein (mg/g tissue)	Lysyl oxidase (SFU)
Wounded control	13.42 ± 4.12	11.39 ± 2.47	11.03 ± 3.19	44.68 ± 3.70	1714 ± 59
Diabetic Induced	10.28 ± 2.20 ^a	8.1 ± 1.30 ^a	8.5 ± 1.42 ^a	26.5 ± 2.40 ^a	1128 ± 37 ^a
<i>C.officinalis</i>	14.72 ± 4.12 ^a	13.49 ± 2.47 ^a	14.01 ± 3.19 ^a	43.48 ± 3.70 ^a	1910 ± 68 ^a

Values are mean ± SD of 6 replications. (SFU- Spectroflourimetric units),p values: ^a:<0.01 vs control.

DISCUSSION

The levels of hydroxyproline and hexosamine, as well as the tissue protein content of the granulation tissue of the dead space injured animals, are shown in Table 2. When normal control injured rats were compared to positive control rats, the amounts of hydroxyl proline, hexosamine, and tissue protein were considerably lower. These levels were raised after treatment with *C.officinalis*. Collagen is a key extracellular matrix protein that helps for wound strength. Collagen not only gives the tissue matrix strength and stability, but it also aids in wound healing by promoting homeostasis and epithelialization.¹⁵ Hydroxyproline is a rare amino acid found in granulation tissue collagen fibres. After applying herbal ointment topically, researchers discovered a rise in hydroxyproline concentration, which indicates enhanced cellular proliferation and hence higher collagen production. Hexosamine and hexuronic acid are matrix molecules that serve as the starting point for the production of new extracellular matrix. Collagen

fibres are known to be stabilised by glycosaminoglycans, which improve electrostatic and ionic interactions with them and may influence their eventual alignment and size. They've been discovered as key determinants of cellular response in development, homeostasis, and illnesses because of their capacity to bind and modify protein interactions.¹⁶

Hexosamine concentrations were substantially higher in the herbal ointment treated groups than in the excision wound control groups in the current investigation, indicating that collagen fibres were stabilised.¹⁷ As a result, damaged tissue with increased hydroxyproline and hexosamine production has the ability to heal. Protein is required for wound healing and granulation tissue formation.

The low protein concentration in excision injured controls indicates that wound healing is delayed due to a prolonged inflammatory phase, fiber plasia

inhibition, and remodelling phase. The animals treated with herbal ointment had a simultaneous rise in total protein content, indicating active synthesis and deposition of matrix proteins in the granulation tissues, which improved wound healing.¹⁸ Streptozotocin has been frequently used to cause diabetes in a number of animals by causing pancreatic β -cell degeneration and necrosis. Similarly, the current investigation comprised STZ induced diabetes followed by an assessment of wound healing capacity utilising a dead space wound model.

Conclusion: Finally, it is reasonable to infer that *C.officinalis* has considerable wound healing activity as a consequence of enhanced collagen

production, wound contraction, and biochemical marker changes, based on the findings of this study. In the future, *C.officinalis* isolated components will be utilised in wound models.

Acknowledgments: The authors are grateful to Deanship of Scientific Research, King Khalid University for sponsoring this study through the Large Research Group Project under grant number RGP 2/186/42.

Conflicts of Interest: “The authors state that they have no competing interests. The funders had no involvement in the study's design, data collection, analysis, or interpretation, manuscript preparation, or the decision to publish the findings.”

REFERENCES

1. Gunasekaran, S., Nayagam, A.A.J. & Natarajan, R. Wound healing potentials of herbal ointment containing *C.officinalis* . on the alteration of immunological markers and biochemical parameters in excision wounded animals. *Clin Phytosci.* 2020;6:77.
2. B.Somashekar Shetty. Analysis of dead space wound granulation tissue in streptozotocin induced diabetic rats. *BCAIJ.*2008;2(2-3):57-59.
3. Tsuchiya H, Sato M, Miyazaki T, Fujiwara S, Tanigaki S, Ohyama M, Tnanka T, linuma M: Comparative study on the antibacterial activity of phytochemical flavanones against methicillin resistant *Staphylococcus aureus*. *J Ethnopharmacol.* 1996;50:27-34.
4. Kishimoto S, Maoka T, Sumitomo K, Ohmiya A. Analysis of carotenoid composition in petals of *Calendula (Calendula officinalis L.)*. *Biosci Biotechnol Biochem.* 2005;69:2122–8.
5. Re TA, Mooney D, Antignac E, Dufour E, Bark I, Srinivasan V, Nohynek G. Application of the threshold toxicological concern approach for the safety evaluation of *Calendula flower (Calendula officinalis L.)* petals and extracts used in cosmetic and personal care products. *Food Chem Toxicol.* 2009;47:1246–54.
6. Fischhof PK, Moslinger Gehmayr R, Herrmann WM, Friedmann A, Russmann DL: Therapeutic efficacy of Vincamine in dementia. *Neuropsychobiology.*1996;34 (1):1345.
7. Chattopadhyay RR, Sarkar SK, Ganguli S, Banerjee RN, Basu TK: Hypoglycemic and antihyperglycemic effect of leaves of *Vinca rosea* Linn. *Indian J Physiol Pharmacol.* 1991;35:145-151.
8. Miliauskas G, Venskutonis PR, Van Beek TA. Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food Chem.* 2004;85:231–7.
9. Muley BP, Khadabadi SS, Banarase NB. Phytochemical constituents and pharmacological activities of *Calendula officinalis L.* (Asteraceae): a review *trop J. Pharm Res.* 2009;8:455–65.
10. Beckmann KH et al. Low level laser therapy for the treatment of diabetic foot ulcers: a critical survey. *Evid Based Complement Alternat Med.* 2014;20(14):1–9.
11. Rosado P et al. Influence of diabetes mellitus on postoperative complications and failure in head and neck free flap reconstruction: a systematic review and meta-analysis. *Head Neck.* 2014;37:615–648.
12. Bagdas D et al. In vivo systemic chlorogenic acid therapy under diabetic conditions: wound healing effects and cytotoxicity/genotoxicity profile. *Food Chem Toxicol.* 2015;81:54–61.
13. Rathi B, Patil PA, Baheti AM: Evaluation of aqueous extract and seeds of *Moringa oleifera* for wound healing in albino rats. *J Nat Remedies.* 2004;4(2):145–149.
14. Komal M Parmar, Priyanka R Shende, Nitin Katare, Mahaveer Dhobi, Satyendra K Prasad, Wound healing potential of *Solanum xanthocarpum* in streptozotocin-induced diabetic rats, *Journal of Pharmacy and Pharmacology.* 2018;70(10):1389–1400.
15. Landsman A, Taft D, Riemer K. The role of collagen bioscaffolds, foamed collagen, and living skin equivalents in wound healing. *Clin Podiatr Med Surg.* 2009;26:525–33.
16. Trownbridge JM, Gallo RL. Dermatan sulfate: new functions from an old glycosaminoglycan. *Glycobiology.* 2002;12(9):117–25.
17. Ricard-Blum S, Ruggiero F. The collagen superfamily: from the extracellular matrix to the cell membrane. *Pathol Biol.* 2005;53:430–42.
18. MacKay D, Meller AL. Nutritional support for wound healing. *Altern Med Rev.* 2003;8(4):359–77