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Anticancer effects of cimetidine

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ABSTRACT

Cancer continues to be a major health problem for those in developed countries being a leading cause of death worldwide. Chemotherapy is able to kill some cancer cells especially the more rapidly replicating tumor cells, but they were nonspecific, characterized by low therapeutic index and associated with a wide range of side effects. Therefore the anticancer field still searching for treatments to avoid these side effects. The in vitro method was used to investigate the effect of pure cimetidine on four types of tumor cell lines [HeLa (human cervical cancer cell line, Passages 18-25), Rhabdomyosarcoma (RD, at 75 passages), Ahmad-Majeed-Glioblastoma-Multiform-2005 (AMGM-5, human cerebral glioblastoma multiform at passages 75-84), Ahmed-Mohammed-Nahi-2003 (AMN-3, spontaneous mammary adenocarcinoma at 158 passages) and normal cell line Rat Embryo Fibroblast (REF, at 87 passages)] in different concentrations and at different exposure times by MTT assay. The results showed that cimetidine exerted significant cytotoxic effects with all concentrations used (31.25-1000 μ g) on all types of cell lines. Because of cytotoxic activity, good pharmacokinetic characteristics and the safety of drug which used for many years in the treatment of peptic ulcer disease, we can conclude that these characteristics make cimetidine a valuable treatment for many types of cancer.

Key Words: Cimetidine, Anticancer, Cell Line, Cervical Carcinoma, Rhabdomyosarcoma, Cerebral Glioblastoma, Mammary Adenocarcinoma

INTRODUCTION

Cancer continues to be a major health problem for those in developed countries being a leading cause of death worldwide and accounting for 7.9 million deaths in 2007. That number is slated to increase to 11.5 million by the year 2030. Lung, stomach, liver, colon and breast cancer cause the most cancer deaths each year [1]. Cancer is a complex set of more than 200 diseases with many causes and stages and histological grades of multiple malignancy [2-4]. This disease takes life's thousands of people of different age and sex every year in Iraq [5]. It has been estimated that approximately 15000 people have been died of cancer in 2005 in Iraq, this number represents 22.8% of the total deaths, moreover, it is expected that such percentage can be increased up to 35.4% in 2030 [6]. Cancer treatments continue to represent a major challenge to medical research [7]. Traditional therapies of cancer (surgery, radiation therapy, and chemotherapy) brought a limited success in treating this disease [8]. Chemotherapy

is able to kill some cancer cells especially the more rapidly replicating tumor cells, but they were nonspecific, characterized by low therapeutic index and associated with a wide range of side effects [9-10]. Therefore the anticancer field still searching for treatments to avoid these side effects [11]. Cimetidine, the first histamine type 2 receptor antagonist to be used clinically, is commonly prescribed to treat gastro esophageal reflux disease, gastric and duodenal ulcers [12]. It has been reported that cimetidine improves the survival of patients with malignant tumors [13-14], including gastric [15], and colorectal carcinomas [16]. The mechanisms involved are incompletely understood. Previous studies showed that cimetidine stimulated the immune response, inhibited the adhesion, invasiveness and metastasis of cancer [13-16], however, the current study was carried out to investigate, if cimetidine exerts direct cytotoxic effect. The direct cytotoxicity of cimetidine in addition to its previously recorded effects will give it an additional value in cancer therapy.

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MATERIALS AND METHODS

The in vitro method was used to investigate the growth inhibitory effect of pure cimetidine on four types of tumor cell lines [HeLa (human cervical cancer cell line at 18-25 passages), Rhabdomyosarcoma (RD, at 75 passages), Ahmad-Majeed-Glioblastoma-Multiform-2005 (AMGM-5, human cerebral glioblastoma multiform at passages 75-84) Ahmed-Mohammed-Nahi-2003 (AMN-3, spontaneous mammary adenocarcinoma at passages 158) and normal cell line Rat Embryo Fibroblast (REF, at passages 87)], in different concentrations and at different exposure times by MTT assay. These cell lines were kindly supplied by experimental therapy department, tissue culture unit/ Iragi centre for cancer and medical genetics research (ICCMGR) maintained in RPMI- 1640 with 10% FCS and MEM with 10% FCS. Cell lines used in this study were subcultured when the cells in the flask formed confluent monolayer, using the previously described protocol [17-18]. The cell viability was determined before studying the cytotoxic effect of the drug on each cell line. Seeding of tryptinized and suspended cells for any cell line in a microtiter plate should be in the range of $(10^4 - 10^5)$ cell /well for the growth cytotoxic assay [19]. Viable cell counting for the study cells were accomplished using trypan blue exclusion. Dead cells take up the dye within a second making them easily distinguishable under the microscope from viable cells which remain unstained. The following protocol was conducted [20].

- a) Cell suspension was prepared (HeLa, AMN3, AMGM and RD cancer cell, REF normal cell).
- b) Clean hemocytometer with its cover slip fixed on its place, was prepared.
- c) One part of cell suspension (0.2 ml) to one part of trypan blue (0.2 ml) to eight parts of PBS (1.6 ml) was mixed. Then 20 µl samples were transferred to the edge of the cover slip, allowed to run into the counting chamber.
- d) After 1-2 minutes counting started with light microscope under 40X objective lens. Separate counts for viable and nonviable cells were recorded.
- e) Cell concentration (cell/ml), total cell count, and cell viability (%) were calculated according to the following equations.

1) C= n \times d \times 10000

Where C= Cell concentration (cell/ml), n= number of counted cells, d= dilution factor=10

2) Total cell count = C (cell/ml) \times the original volume of fluid from which the cell sample was taken.

3) Cell viability (%) = total viable cells (unstaind) total cells counted (staind & unstaind) × 100

Cytotoxicity assay:

Preparation of drugs stock solution: Pure cimetidine was obtained from state company for drug industries & medical appliances (SDI) - Samarra /Iraq. Stock solutions of this drug were prepared for cytotoxic assay (cell growth inhibition assay), by dissolved 0.01 g of cimetidine in 1ml triple distal water and filtered by 0.22µm syringe filter.

Preparation of cell lines for cytotoxic assay: Cell cultures in microtiteration plate (96 wells) were exposed to cimetidine at six concentrations during the log phase of growth and the effect was determined after the end of exposure time. The following method was used for cytotoxic assay:

a. Seeding: After cells in the incubated falcon became monolayer, the confluent monolayer was trypsinzed, then 200 μ l of cell suspension seeds in microtitration plates were dispensed into each well, except wells at edges of plate to reduce the edge effect, that every well contain about $10^4 - 10^5$ cells/well and then coved by plate lids and sealed with self adhesive film then shacked gently and returned to the incubator.

b. Incubation: Microtitration plates were then incubated at 37°c until the cells reached confluence (i.e., vary according to the types of cell line). After cells attachment, the plate was checked out for contamination.

c. Exposure: When the cells are in the exponential phase exactly in population doubling time (PTD), which the cells in full of its activity (depending on the growth curve of each cell lines), cells were exposed to six concentration of cimetidine (1000, 500, 250, 125, 62.5 and 31.25 μ g/ml) (Four replicates for each tested concentration). 200 μ l of maintenance medium added to each well of control group (twelve wells were used).

d. Staining: Cell viability was measured after 24, 48 and 72 hrs of exposure by removing the medium, adding 28 μ l of 2 mg/ml solution of MTT and incubating for 1.5 hrs at 37°C. After removing the MTT solution, the crystals remaining in the wells were solubilised by the addition of 130 μ l of X 100

Dimethyl Sulphoxide (DMSO) followed by 37°C incubation for 15 min with shaking.

The absorbency was determined on a microplate reader at 550 nm (test wavelength); the assay was performed in triplicate [21]. The inhibiting rate of cell growth was calculated as follow [22]: Inhibition rate =

mean of control-mean of treatment

mean of control

RESULTS

The results showed that cimetidine decreased the growth of AMGM5 cells significantly as compared to untreated control cells; it appeared that the growth inhibition was concentration and exposure time dependent. The results showed that the inhibition rates for the concentrations 31.25. 62.5, 125, 250, 500 and 1000μ g/ml were 4.900, 11.342, 195.512, 27.125, 34.295 and 42.754% respectively after 24 hrs of exposure. When the exposure time increased to 48 hrs, the inhibition rates for these concentrations reached 15.497, 24.931, 31.912, 42.319, 50.753 and 58.587 % respectively. However after 72 hrs exposure the inhibition rates increased to 22.181, 32.617, 40.087, 48.694, 60.076 and 67.647% respectively (table 1).

As shown in the table 2, cimetidine decreased the growth of AMN3 cells significantly as compared to untreated control cells; the growth inhibition was also concentration and exposure time dependent. The results showed that the inhibition rates for the concentrations 31.25. 62.5, 125, 250, 500 and 1000µg/ml were 8.713, 16.819, 23.106, 28.910, 34.278 and 42.365% respectively after 24 hrs of exposure. After 48 hrs of exposure the inhibition rates increased to18.497, 24.126, 31.007, 37.981, 44.553 and 48.976% respectively. When the exposure period increased to 72 hrs, the inhibition rates reached 21.637, 33.137, 42.186, 50.163, 58.352 and 65.243% respectively.

Cimetidine also significantly decreased the growth of HeLa cells in comparison to untreated control cells with a concentration and exposure time dependent manner. When the HeLa cell line exposed to 31.25. 62.5, 125, 250, 500 and 1000µg/ml concentrations of cimetidine, the growth rates inhibited by 7.145, 12.567, 16.872, 24.506, 32.527 and 39.845% respectively after 24 hrs exposure. The same concentrations of the drugs exerted inhibition of growth rates 18.084, 25.251, 30.257, 40.595, 46.988 and 54.658%, when the exposure time increased to 48 hrs. However, after 72 hrs of exposure, the growth rates inhibition reached 31.740, 39.127, 48.080, 57.751, 65.141 and 73.060% for the same concentrations respectively (table 3).

The results also showed that cimetidine decreased the growth of RD cells significantly as compared to untreated control cells with a concentration and exposure time dependent manner. The inhibition rates for the concentrations 31.25. 62.5, 125, 250, 500 and 1000 μ g/ml were 7.145, 12.567, 16.872, 24.506, 32.527 and 39.845% respectively after 24 hrs of exposure. When the exposure time increased to 48 hrs, the inhibition rates for these concentrations reached 18.084, 25.251, 30.257, 40.595, 46.988 and 54.658% respectively. After 72 hrs exposure the inhibition rates increased to 31.740, 39.127, 48.080, 57.751, 65.141 and 73.060 % respectively (table 4).

Against normal cell line rat embryo fibroblast (REF), cimetidine in concentration of 31.25. 62.5, 125, 250, 500 and 1000μ g/ml also exerted significant growth inhibition rates (5.132, 11.704, 16.931, 20.520, 26.207 and 30.908 % respectively) after 72 hrs exposure (table 5).

DISCUSSION

It has been reported that cimetidine improves the survival of patients with malignant tumors, including gastric and colorectal carcinomas [14-16, 23-24]. The mechanisms by which cimetidine improve survival rate in gastrointestinal tumors were included enhancement of the host immune response against tumor cells via blocking of histamine H2 receptors. By this mechanism, it enhanced infiltrating of lymphocytes in the tumors [16, 25-27]. Cimetidine also exerted an inhibitory effect on cancer cell migration and adhesion to endothelial cells, thus inhibiting tumor metastasis [28-32]. It was also reported that cimetidine inhibited colon adenocarcinoma cell adhesion to vascular endothelial cells and prevents metastasis by blocking E-selectin expression [31-32]. Eselectin is able to bind to certain types of complex carbohydrate chains that are frequently expressed on the surfaces of cancer cells but not by the healthy tissues from which they arise. Otherwise, cimetidine makes endothelial cells more slippery by suppressing the E-selectin adhesion protein, thus making it harder for cancer cells circulating in the bloodstream to bind to the endothelial lining of blood vessels [33-34]. However, in addition to the previously mentioned beneficial effects, this study also showed that cimetidine exerted direct cytotoxic effects on cell lines, this direct effect could be attributed to blocking of H2 receptors (H2R). Histamine regulates diverse biological responses related to tumor growth including proliferation, differentiation and apoptosis, which

indicate that histamine is a crucial mediator in cancer development and progression. [35-39]. So, overexpression of histidine decarboxylase (HDC), (the only enzyme responsible for the generation of histamine from L-histidine) at both the mRNA and protein levels, with an increased levels of histamine have been recorded in melanoma, small cell lung carcinoma, breast carcinoma, endometrial cancer and colorectal carcinoma [40-45]. In the other hand, inhibition of HDC with monofluormethyl histidine resulted in antitumoural effects on experimental tumors in rodents. Furthermore, the employment of specific HDC antisense oligonucleotides suppressed melanoma cell proliferation [42, 45-48]. Therefore the cytotoxicity of cimetidine could be related to

blocking of H2R of histamine which promote tumor proliferation. Cimetidine has been in use for a number of years. It is generally well tolerated, and appears to be quite safe. Furthermore the drug characterized by a good pharmacokinetic characteristics, so, the effect of six months therapy with cimetidine (800 mg or 1600 mg/day) showed that the mean elimination half-life of cimetidine was 100±25 min, the total body cimetidine clearance was 652 + 223 ml/min, the mean volume of distribution at steady state was 65 ± 181 and the overall bioavailability was 78% [49-50]. Accordingly, we can conclude that the good pharmacokinetic characteristics, safety and direct broad anticancer effects make cimetidine a valuable additional treatment for many types of cancer.

Table 1: Growth inhibitory rate of different concentrations of cimetidine on AMGM5 cell line after 24, 48 and 72 hrs of exposure.

| Conc. µg | Effects according to the period of exposure | | | | | | |
|----------|---|------------------------------|------------------------------|-------------------------------|------------------------------|------------------------------|--|
| | 24 hrs | | 48 hrs | | 72 hrs | | |
| | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | |
| Control | 0.605±0.030 | | 0.606±0.015 | | 0.594 ±0.020 | | |
| 31.25 | 0.565±0.040* | 4.900±1.194 | 0.480±0.032** | 15.497±2.655 | 0.471±0.047** | 22.181±3.087 | |
| 62.5 | 0.518±0.053** | 11.342±0.331 | 0.426±0.029*** | 24.931±2.816 | 0.408±0.036**** | 32.617±1.992 | |
| 125 | 0.478±0.035*** | 19.512±1.504 | 0.386±0.030**** | 31.912±2.085 | 0.362±0.030**** | 40.087±1.054 | |
| 250 | 0.434±0.049**** | 27.125±2.881 | 0.328±0.035**** | 42.319±4.276 | 0.309±0.017**** | 48.694±2.873 | |
| 500 | 0.391±0.049**** | 34.295±2.929 | 0.279±0.027***** | 50.753±3.194 | 0.240±0.018***** | 60.076±3.988 | |
| 1000 | 0.341±0.043**** | 42.754±3.041 | 0.234±0.014***** | 58.587±2.503 | 0.194±0.024***** | 67.647±5.433 | |

In comparison with control , * (P < 0.05), ** (P < 0.01), ***(P < 0.001), ****(P < 0.0001), ****(P < 0.0001).

| Con. µg | Effects according to the period of exposure | | | | | | |
|------------|---|------------------------------|------------------------------|-------------------------------|------------------------------|------------------------------|--|
| | 24 hrs | | 48 hrs | | 72 hrs | | |
| | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | |
| Control | 1.053 ± 0.088 | | 1.053± 0.073 | | 1.105±0.074 | | |
| 31.25 | 0.904±0.046* | 7.145±6.832 | 0.879±0.0974** | 18.084±4.604 | 0.727±0.141**** | 31.740±11.832 | |
| 62.5 | 0.846±0.042** | 12.567±7.184 | 0.790±0.090*** | 25.251±1.802 | 0.649±0.168**** | 39.127±15.950 | |
| 125 | 0.804±0.041*** | 16.872±6.641 | 0. 54±0.137**** | 30.257±3.458 | 0.553±0.144**** | 48.080±13.939 | |
| 250 | 0.731±0.033*** | 24.506±5.599 | 0.643±0.139**** | 40.595±4.691 | 0.449±0.095***** | 57.751±7.786 | |
| 500 | 0.654±0.042**** | 32.527±4.229 | 0.578±0.147**** | 46.988±8.165 | 0.370±0.080***** | 65.141±6.575 | |
| 1000 | 0.584±0.074**** | 39.845±7.448 | 0.498±0.155***** | 54.658±10.386 | 0.286±0.064***** | 73.060±6.456 | |

Table 2: Growth inhibitory rate of different concentrations of cimetidine on AMN3 cell line after 24, 48 and 72 hrs of exposure.

In comparison with control, * (P<0.05), ** (P<0.01), ***(P<0.001), ****(P<0.0001), ****(P<0.0001).

Table 3: Growth inhibitory rate of different concentrations of cimetidine on HeLa cell line after 24, 48 and 72 hrs of exposure.

| Conc. µg | Effects according to the period of exposure | | | | | | |
|----------|---|------------------------------|------------------------------|-------------------------------|------------------------------|------------------------------|--|
| | 24 hrs | | 48 hrs | | 72 hrs | | |
| | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | |
| Control | 0.685±0.075 | | 0.610±0.014 | | 0.657±0.066 | | |
| 31.25 | 0.584±0.095* | 8.713±9.180 | 0.480±0.032** | 18.497±11.631 | 0.471±0.047*** | 21.637±9.712 | |
| 62.5 | 0.532±0.084** | 16.819±8.036 | 0.426±0.029*** | 24.126±3.696 | 0.408±0.036**** | 33.137±4.083 | |
| 125 | 0.491±0.072*** | 23.106±6.037 | 0.386±0.030**** | 31.007±2.621 | 0.362±0.030**** | 42.186±1.843 | |
| 250 | 0.453±0.056*** | 28.910±2.591 | 0.328±0.035**** | 37.981±1.085 | 0.309±0.017***** | 50.163±1.306 | |
| 500 | 0.436±0.050**** | 34.278±2.988 | 0.279±0.027**** | 44.553±1.176 | 0.240±0.018***** | 58.352±2.365 | |
| 1000 | 0.368±0.059**** | 42.365±3.963 | 0.234±0.014**** | 48.976±3.281 | 0.194±0.024**** | 65.243±5.758 | |

| Conc. µg | Effects according to the period of exposure | | | | | | |
|-------------|---|------------------------------|------------------------------|-------------------------------|------------------------------|------------------------------|--|
| | 24 hrs | | 48 hrs | | 72 hrs | | |
| | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | Optical density (mean±SD) | Inhibition rate (mean±SD) | |
| Control | 0.588±0.014 | | 0.593±0.017 | | 0.588±0.008 | | |
| 31.25 | 0.551±0.061* | 5.957±5.638 | 0.553±0.040* | 8.399±5.761 | 0.477±0.033** | 21.622±5.555 | |
| 62.5 | 0.527±0.054* | 8.996±7.899 | 0.509±0.030** | 15.664±3.721 | 0.434±0.030*** | 28.638±5.623 | |
| 125 | 0.491±0.063** | 15.363±10.243 | 0.471±0.048*** | 21.869±10.038 | 0.393±0.029**** | 35.450±4.635 | |
| 250 | 0.445±0.053** | 23.264±7.984 | 0.423±0.058**** | 29.869±10.038 | 0.337±0.024**** | 44.660±4.804 | |
| 500 | 0.412±0.060*** | 28.972±9.395 | 0.388±0.056**** | 35.766±9.690 | 0.304±0.019***** | 50.065±2.584 | |
| 1000 | 0.358±0.063**** | 38.155±6.933 | 0.348±0.051**** | 42.295±8.873 | 0.260±0.033***** | 57.932±4.957 | |

Table 4: Growth inhibitory rate of different concentrations of cimetidine on RD cell line after 24, 48 and 72 hrs of exposure.

In comparison with control , * (P<0.05), ** (P<0.01), ***(P<0.001), ****(P<0.0001), *****(P<0.0001).

Table 5: Growth inhibitory rate of different concentrations of cimetidine on REF cell line after 72 hrs of exposure.

| Concentration µg | Effect after exposure for 72 hrs | | | |
|------------------|----------------------------------|---------------------------|--|--|
| | Optical density (mean±SD) | Inhibition rate (mean±SD) | | |
| Control | 0.859±0.028 | | | |
| 31.25 | 0.790±0.073* | 5.132±1.675 | | |
| 62.5 | 0.735±0.064** | 11.704±2.916 | | |
| 125 | 0.692±0.068** | 16.931±3.465 | | |
| 250 | 0.663±0.070*** | 20.520±3.701 | | |
| 500 | 0.610±0.081*** | 26.207±5.213 | | |
| 1000 | 0.578±0.092**** | 30.908±6.973 | | |

In comparison with control, * (P<0.05), ** (P<0.01), *** (P<0.001), **** (P<0.0001).

REFERENCES

- 1. American Association for Cancer Research. AACR Cancer Progress Report 2013. Clin Cancer Res 2013; 19(Supplement 1): S1-S88.
- 2. Augenlicht LH. Chemoprevention : intermediate markers In Encyclopedia of cancer, Vol 1. Academic press New York 1997.
- 3. Parkin DM et al . Global cancer statistics 2002. Cancer J Clin 2005; 55: 74–108.
- 4. Mathers CD, Loncar D. Projections of global mortality and burden of disease from 2002 to 2030. PLoS Med 2006; 3: e442.
- 5. Ministry of Health. Results of Iraqi Cancer Registry 1999-2001, Iraqi Cancer Board, Baghdad, Iraq 2009.
- 6. World Health Organization. Cancer 2005.
- 7. Jemal A et al. Cancer statistics, 2007. Cancer J Clin 2007; 57:43-66.

- 8. Kintzios SE. What do we know about cancer and its therapy? In: Plants That Fights Cancer. Kintzios SE and Barberaki MG (Eds). CRC Press, USA 2004: 1-14.
- 9. Al- Snafi AE. Antimicrobial drugs. Al Diaa Publication house, Iraq 2013: 101-14.
- 10. Al- Snafi AE. Pharmacology and therapeutics. Al Diaa Publication house, Iraq 2013: 503-16.
- 11. Inche AG, Thangue NBL. Chromatin control and cancer-drug discovery: realizing the promise. Drug Discovery Today 2006; 11: 97-109.
- 12. Freston JW. Cimetidine I. Developments, pharmacology, and efficacy. Ann Intern Med 1982; 97: 573-80.
- 13. Burtin C et al. Clinical improvement in advanced cancer disease after treatment combining histamine and H2-anti-histaminics (ranitidine or cimetidine). Eur J Cancer Clin Oncol 1988; 24:161-67.
- 14. Siegers CP et al. Does cimetidine improve prospects for cancer patients? A reappraisal of the evidence to date. Digestion 1999; 60: 415-421.
- 15. Tønnesen H et al . Effect of cimetidine on survival after gastric cancer. Lancet 1988; 2: 990-92.
- Adams WJ et al. Cimetidine inhibits in vivo growth of human colon cancer and reverses histamine stimulated in vivo growth. Gut 1994; 35:1632-36.
- 17. Freshney RS. Culture of animal cells. A manual of Basic Technique, 3rd ed. New York Press 1994 : 287.
- 18. Neri D et al. Protocols for the preparation of tumour cells for sc injections in mice. Tumor Immunol 2012; 12(3):16.
- 19. Freshney I. Culture of animal cells. A manual basic technique, 4th edition. Wiley-Liss 2000.
- 20. Darling DC, Morgan SJ. Animal cells : Culture and Media, Essential Data. John Wiley and Sons, Chichester 1994:90-116.
- Betancur-Galvis L et al. Cytotoxic and antiviral activity of Colombian medicinal plant extract of the Euphorbia genus. Mem Inst Oswaldo Cruz Rio de Janeiro 2002; 97(4): 541-6.
- Betancur-Galvis L et al. Antitumor and antiviral activity of Colombian medicinal plant extracts. Mem Inst Oswalo Cruz, Rio de Janeiro 1999; 94(4):531-35.
- Kelly MD et al. Randomized trial of preoperative cimetidine in patients with colorectal carcinoma with quantitative assessment of tumor-associated lymphocytes. Cancer 1999; 85(8): 1658-63.
- 24. Matsumoto S et al. Cimetidine increases survival of colorectal cancer patients with high levels of sialyl Lewis-X and sialyl Lewis-A epitope expression on tumour cells. British Journal of Cancer 2002; 86: 161–7.
- Adams WJ, Morris DL. Pilot study cimetidine enhances lymphocyte infiltration of human colorectal carcinoma. Cancer 1997; 80: 15–21.
- 26. Hansbrough JF et al. Prevention of alterations in postoperative lymphocyte subpopulations by cimetidine and ibuprofen. Am J Surg 1986; 151:249-55.
- Adams WJ et al. Cimetidine preserves non-specific immune function after colonic resection for cancer. Aust N Z J Surg 1994; 64: 847-52.
- 28. Sasson AR et al.Cimetidine: an inhibitor or promoter of tumor growth. Int J Cancer 1999; 81: 835-8.
- 29. Lawson JA et al. Ranitidine and cimetidine differ in their in vitro and in vivo effects on human colonic cancer growth. Br J Cancer 1996; 73: 872-876.
- 30. Lefranc F et al. Cimetidine, an unexpected anti-tumor agent, and its potential for the treatment of glioblastoma. Int J Oncol 2006; 28: 1021-1030..
- 31. Kobayashi K et al. Cimetidine inhibits cancer cell adhesion to endothelial cells and prevents metastasis by blocking E-selectin expression. Cancer Res 2000; 60: 3978-84.
- 32. Fukuda M et al. Cimetidine induces apoptosis of human salivary gland tumor cells. Oncol Rep 2007; 17(3):673-8.
- 33. Liu FR et al. Cimetidine inhibits the adhesion of gastric cancer cells expressing high levels of sialyl Lewis x in human vascular endothelial cells by blocking E-selectin expression. Int J Mol Med 2011; 27(4): 537-44.
- 34. Kannagi R et al. Carbohydrate-mediated cell adhesion in cancer metastasis and angiogenesis. Cancer Sci 2004; 95(5): 377-84.
- 35. Leurs R et al. Molecular pharmacological aspects of histamine receptors. Pharmacology & Therapeutics 1995; 66 (3): 413-63.
- 36. Medina VA, Rivera ES. Histamine receptors and cancer pharmacology. Br J Pharmacol 2010; 161(4)755-67.
- 37. Davio CA et al. H1 and H2 histamine receptors in experimental carcinomas with an atypical coupling to signal transducers. Biochemical Pharmacology 1995; 50 (1): 91-6.
- 38. Fukushima Y et al. Oligomer formation of histamine H2 receptors expressed in Sf9 and COS7 cells. FEBS letters 1997; 409 (2): 283-6.
- Fitzsimons C et al. Regulation of phospholipase C activation by the number of H(2) receptors during Ca(2+)-induced differentiation of mouse keratinocytes. Biochemical Pharmacology 2002; 63(10): 1785-96.
- Hegyesi H et al. Suppression of melanoma cell proliferation by histidine decarboxylase specific antisense oligonucleotides. J Invest Dermatol 2001; 117:151-3.
- Graff L et al. Expression of histidine decarboxylase and synthesis of histamine by human small cell lung carcinoma. Am J Pathol 2002; 160: 1561-5.
- 42. Garcia-Caballero M et al. Histamine synthesis and content in benign and malignant breast tumours. Surg Oncol 1994; 3:167-73.
- 43. Chanda R, Ganguly AK. Diamine-oxidase activity and tissue di- and poly-amine contents of human ovarian, cervical and endometrial carcinoma. Cancer Lett 2001; 89: 23-8.
- 44. Garcia-Caballero M et al. Increased histidine decarboxylase (HDC) activity in human colorectal cancer: results of a study on ten patients. Agents Actions 1988; 23: 357-60.
- 45. Bartholeyns J et al. Involvement of histamine in growth of mouse and rat tumors: antitumoral properties of monofluormethyl histidine, an enzyme-activated irreversible inhibitor of histidine decarbolxylase. Cancer Res 1984; 44: 639–45.
- 46. Falus A et al. Paracrine and autocrine interactions in melanoma: histamine is a relevant player in local regulation. Trends Immunol 2001; 22: 648–52.
- 47. Pós Z et al. Histamine and cell proliferation. In: Histamine: Biology and Medical Aspects. Falus A (Editor). Budapest, Spring Med Publishing 2004: 199–217.
- 48. Garcia Rodriguez LA, Jick H. Risk of gynaecomastia associated with cimetidine, omeprazole, and other antiulcer drugs. BMJ 1994; 308(6927): 503-6.
- 49. Webster J et al. Cimetidine a clinical and pharmacokinetic study. Br J Clin Pharmac 1981; 11: 333-8.