



Antimicrobial Activity of Natural Edible Gums

Bhoj Raj Singh*¹, Sakshi Dubey¹, Mahtab Zakira Siddiqui²

¹Division of Epidemiology, ICAR-Indian Veterinary Research Institute, Izatnagar-243122, India

²P.P.D. Division, ICAR-Indian Institute of Natural Resins and Gums, Namkum, Ranchi-834 010, India

Received: 27-09-2015 / Revised: 20-10-2015 / Accepted: 25-10-2015

ABSTRACT

Edible gums are tears or flakes of dried sap (exudates) of many (mostly thorny) trees and shrubs either after natural or manmade injuries to wooden stem and branches. They are ionic, natural, complex polymers of glycopeptides containing several other compounds and mineral salts. Many of the gums are edible specifically those obtained from trees/ shrubs of Fabaceae family including *Acacia*, *Sterculia*, *Astragalus*, *Balanites*, *Buchnania* and *Anogeissus* species. Gums have been used since centuries for their nutritional, culinary and therapeutic properties in food, pharmacy and refining industries. In recent past gums have become very important starting material for green synthesis of nanomaterials including metallic (silver, copper, palladium etc.) nanoparticles and nanotubes due to their reductant and stabilizer activity. Their biogenic synthesis utility is mainly interesting due to their biodegradability, nontoxic, non-mutagenic nature, natural availability, higher resistance to microbial attacks and long shelf-life. Many of the gums are known to alleviate infections like diarrhoea, dysentery, sore-throat, kidney, wound and gum infections. However, only scattered reports are in literature evaluating their antimicrobial potential leading to underestimation and sometime over expectations from gums and gum-products in therapeutics. This short review is for understanding the available information on edible gums as herbal antimicrobial agents.

Key words: Gondh, Gond, Antibacterial, Anti-diarrhoeal, Green synthesis, Nanoparticles, Gum Arabic, Gum Ghatti, Gum Tragacanth, Gum Karaya.



INTRODUCTION

Most of the edibles gum from trees are actually exudates gums and made of sugar polymers. Natural gums often known as Gond, Goond, Dinka, Gaund or Gondh are polysaccharides of plant origin recommended for benefits in pregnancy and after delivery to strengthens back bone. They are water soluble and produce gel like viscous solutions and for this quality have utility in food industry as thickening, gelling, emulsifying and stabilizing agents [1]. For their utility as adhesives, binding agents, crystal inhibitors, clarifying agents, encapsulating agents, flocculating agents, swelling agents and foam stabilizers etc. natural gums have applications in many industries including vineyards as refining agents [2]. Chemical composition of gum varies greatly depending on plant species, agro climatic conditions and time of harvest [3]. Gums are complex mixture of glycoproteins and polysaccharides and natural source of arabinose and ribose. Arabinogalactan, major constituent of gum

arabic and other edible gums is a polymer of arabinose and galactose linked to protein moiety forming glycopeptides. Gums are slightly acidic in nature due to 8-5' non cyclic diferulic acid covalently linked to carbohydrate arm of arabinogalactan-protein [4] and are referred as polyelectrolytes (ionic polymers).

The most widely used edible gum is gum arabic, dried sap (exudates) of *Acacia* spp. trees and is also known as acacia gum, char gund, char goond, or meska [5, 6]. It is mostly obtained from trees of two *Acacia* species viz., *A. senegal* and *A. seyal*. Besides, gum ghatti (from *Anogeissus* trees), gum tragacanth (from *Astragalus* shrubs, gond katira), karaya gum (from *Sterculia* trees), gum babul (from *Acacia nilotica*), gum chironji (from *Buchnania lanzan* trees), and gums from *Balanites aegyptica* and *Prosopis juliflora* are also edible gums and available in market [7].

Gums are used since ages for their nutritional, culinary and therapeutic values throughout the

*Corresponding Author Address: Dr. Bhoj R Singh, Head Division of Epidemiology, Indian Veterinary Research Institute, Izatnagar-243122, India; Email: brs1762@ivri.res.in; brs1762@gmail.com

world. Among the common edible gums in India, gums obtained from *Acacia nilotica* (Babul), *Buchnania lanzan* (Chironji), *Sterculia* (Karaya), *Anogeissus latifolia*, *Balanites aegyptica* and *Prosopis juliflora* [5] are in commonplace [7] and have different utilities in food, pharma and other industries.

Gum Arabic: The most commonly available and most used edible gum all over the globe has multiple uses. *Acacia* gum (gum Arabic) contains chiefly arabin, mixture of calcium, magnesium and potassium salts of arabic acid and also contains enzymes like oxidases, peroxidases and pectinases [8]. Gum arabic is water soluble and can be used as decoctum and syrup in therapeutic uses. In therapeutics gum arabic is known to be beneficial in kidney diseases [5] and is known for its analgesic, astringent, emollient, liver tonic, antipyretic and anti-asthmatic properties [6, 9]. It has been claimed to act as an anti-oxidant, and protects in experimental hepatic, renal and cardiac toxicities in rats [6, 10]. Gum acacia has been used for cure of diarrhoea, dysentery and diabetes [6, 10]. It is used in diarrhoea, dysentery and diabetes, dry cough in amoebic dysentery, as a tonic, anti-asthmatic, analgesic and in oral cavity lesions.

Antimicrobial activity of *Acacia* is reported to be concentrated either in leaves or bark and their extracts have been reported inhibitory against *Streptococcus viridans*, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella* Typhi, *Bacillus subtilis*, *B. creus*, *Shigella sonnei* and even against *Candida albicans*, *C. glabrata* and *Aspergillus niger* and *Rhizoctonia solani* [9, 11-15]. The antimicrobial activity has been chiefly detected in methanolic extracts rather than in aqueous extract [14]. Even the leaf extract was found active against strains of *Serratia marcescens*, *Erwinia herbicola*, *Xanthomonas* sp. *Arthrobacter chlorophenolicus* and fungal strains of *Botrytis cineria*, *Fusarium oxysporum*, *A. flavus* [14]. Antimicrobial activity was detected in ethanolic extract but not in chloroform, benzene, ether or aqueous extracts [15]. Flavonoids and alkaloids from leaves of *Acacia nilotica* inhibited *S. aureus*, *B. subtilis*, *E. coli* and *S. Typhi*, but not inhibited *P. aeruginosa* and *K. pneumoniae*. For sensitive strains, it had minimum inhibitory concentration (MIC) of 0.125 to 1mg/ ml [16]. Besides, saponins of *Acacia* spp. inhibit *P. aeruginosa* and polysaccharides inhibit *S. aureus* and *B. subtilis* [16].

Though antimicrobial activity of gum acacia (gum arabic) is rarely reported [17, 18] it has been successfully used as carrier of other antimicrobial herbal preparations (garlic extract or cinnamon) to form an antimicrobial coating on foods [19].

Besides, metal arabates (mercury, silver and copper) prepared using arabic acid of gum acacia have potent antimicrobial activity against *E. coli*, *B. subtilis*, *B. anthracis*, *B. pumilus*, *S. Typhi*, *S. aureus*, *Streptococcus pyogenes*, *Proteus vulgaris* and *A. niger*, *A. flavus*, *Trichophyton equingia*, *Fusarium oxysporum* and *Cryptococcus neoformans* [20]. Alcoholic extract of gum Arabic from *A. nilotica* has rare report for antimicrobial potential but from *Acacia senegal*, *Acacia seyal* aqueous extracts is active against *S. aureus*, *S. epidermidis*, *St. pneumoniae*, *P. aeruginosa*, *Proteus merabilis*, *Acinetobacter*, *Enterobacter*, *Klebsiella pneumoniae*, *Serratia* spp., *E. coli*, *Salmonella* Typhi and *C. albicans* in agar well diffusion method [17, 21]. Alcoholic extract of gum *Acacia nilotica* has been shown to inhibit *E. coli*, *B. cereus* and *Cerospora pongamae* but not inhibited *A. niger* which could be inhibited by its hot water extract [18].

Gum Chironji: Chironji (*Buchnania lanzan*) gum is light brown or golden coloured amorphous solid. It is known for its several therapeutic uses in Unani and Ayurvedic medicine as analgesic (nerve soothing), anti-rheumatic and antidiarrhoeic agent [7].

Similar to *Acacia*, chironji leaves are known to contain antimicrobial activity [22]. Chironji leaves contain saponins, flavanoids, steroids, cardiac glycosides, carbohydrate, tannins and phenolics. Its alcoholic extract has the maximum activity and minimum activity is detected in aqueous extract. Its extract can inhibit *E. coli*, *P. aeruginosa* and *S. aureus*, *A. niger* and *Penicillium* sp. [22]. Among ethanol and methanol, latter has been found to extract better antimicrobial activity of chironji leaves and extract could inhibit *E. coli*, *S. Typhi*, *S. aureus* and *B. subtilis* [23]. Only little antimicrobial activity is reported in petroleum ether, chloroform and water extracts of *B. lanzan* leaves [22, 23]. There is hardly any reference to antimicrobial activity in gum chironji [7]. Gum chironji is reported beneficial in intercostals' pain and diarrhoea. But more commonly used for adulteration of guggul (oleoresin from *Commiphora wightii*) by adding some perfume/scent [7].

Gum Tragacanth: Obtained from *Astragalus* shrubs and is commonly known as gond katira. The *Astragalus* shrubs, known as locoweeds are native to arid regions of the eastern Mediterranean and South Western Asia [24]. The common gum producing species are *A. gummifer*, *A. adscendens* and *A. microcephalus*. The name "tragacanth" has origin from ribbon shape of exuded gum, similar to goat horn ("tragos")

meaning goat and "akantha" meaning horn) in morphology. Tragacanth gum has been used as laxative, antitussive, anti-diarrhoeic, and as an aphrodisiac. It is used as an adhesive agent in making pills and tablets, as an emulsifying agent in lotions, creams and pastes and as a thickener in cosmetics, toothpastes, jellies, salad dressings, syrups, mayonnaise, sauces, liqueurs, candy, ice cream and popsicles [24].

Antimicrobial activity of ethanolic extract of gum acacia, gum tragacanth and guar gum has been detected against *St. pneumoniae*, *S. aureus*, *E. coli*, *K. pneumoniae* and *P. aeruginosa*, and *C. albicans* [25]. Tragacanth gum extract had significantly higher antimicrobial activity than other gum extracts [25]. Gum tragacanth, similar to Karaya and Ghatti gums, is not generally considered to have any significant antimicrobial activity but has been used as an ideal material for green synthesis of nanoparticles and nano-fiber scaffolds having application in infection control due to their broad spectrum antimicrobial activity [26]. Due to biodegradability, nontoxic, non-mutagenic nature, natural availability, higher resistance to microbial attacks and long shelf-life natural gums are ideal materials for nanosynthesis of microcapsules loaded with herbal extracts [27]. Besides, tragacanth gum fed at a level of 3% along with 3% cholesterol in the diet of cockerels inhibited the development of hypercholesterolemia [28]. Instead of getting inhibited some of the potential pathogens including species of *Klebsiella*, *Serratia*, and *Yersinia* ferment tragacanth gum indicating its rapid biodegradability in intestine [29].

Gum *Balanites aegyptiaca* (Hingot Gum): *Balanites aegyptiaca* (L), soapberry tree or thorn tree or desert date occurs throughout dry and hot regions of Africa and parts of Middle East including India and is known as Hingot tree [30]. The different parts of the plant have utility in therapy of bacterial infections, viral fever and parasitic infestations. It has been used as an antidote for arrow poison and snake venom [30]. Antibacterial and antimycotic activity has been reported in *B. aegyptiaca* leaves, bark and fruit mesocarps [31-34]. Antimicrobial ingredients could be extracted in more concentrated form in aqueous extract than in alcoholic extracts [31]. However, in some studies [35] growth inhibition of metallo-B-lactamase producing *E. coli*, *Klebsiella* spp. and *Citrobacter* spp. was observed both in alcoholic and aqueous extracts of fruit mesocarp of *B. aegyptiaca*. However, very little is understood about edible gum of *B. aegyptiaca* or its antimicrobial activity. Gum mixed with maize meal porridge has been reported useful in treatment of chest complaints [36].

Kikar Gum: It is dried wooden sap of *Prosopis juliflora* (Vilayati kikar) plant. The exudates gum of *P. juliflora* is odourless with little taste. It is a non-crystalline water soluble substance but insoluble in alcohol or ether [37]. It has been used to treat eye infections [37]. Though gum has rarely been studied for its antimicrobial activity, the bark of the tree (source of gum) has been shown to possess antimicrobial activity against *E. coli*, *Proteus mirabilis*, *P. aeruginosa*, *K. pneumoniae*, *S. Paratyphi B*, *Shigella flexneri*, *St. pyogenes* and *S. aureus* in its methanolic extract due to alkaloids in it, the MIC of bark extract was determined in range of 4.6 to 7.5 mg/ml [38]. Hot and cold aqueous extracts of *P. juliflora* leaves is reported as a potent antibacterial at 100 mg/ml concentration against *B. subtilis*, *E. coli*, *E. faecalis*, *K. pneumoniae*, *P. aeruginosa*, *S. aureus*, *S. epidermidis*, *St. pyogenes*, *S. Typhi* and *S. Typhimurium* [39].

Gum Ghatti (Indian Gum, Ghatti Gum): It is a water-soluble complex proteinaceous polysaccharide bark exuded of *Anogeissus latifolia* tree found in dry-deciduous forests of India and Sri Lanka. It is glossy and amorphous, found in rounded tears or in vermiform masses of off white to dark brown colour [40]. It is rich in calcium and magnesium and on hydrolysis produces arabinose, galactose, mannose, xylose and glucuronic acid. It is soluble in water; 1 g in 5 ml water forms viscous adhesive mucilage. It is an alternative hydrocolloid approved for the food industry as a thickener and emulsifying agent [40]. It is also used as an emulsifier and stabilizer in beverages, table syrups, petroleum and in non-petroleum waxes to form liquid and wax paste emulsions and flavor fixative [41]. It is mainly used as thickening agent and stabilizer and is a promising polysaccharide for pharmaceutical purposes [41]. Recently it has been used to formulate responsive hydrogels [42]. Though gum ghatti has no antimicrobial activity it has been used for green (biogenic) production of silver nanoparticles with wide spectrum antimicrobial potential [43]. Gum ghatti has been exploited for its dual functional reductant and stabilizer activity for the synthesis of nanoparticles [43]. Gum ghatti imparted better antibacterial activity to silver nanoparticles synthesized with it (Ag NP-GT) than that could be produced with gum olibanum [43]. It has also been used for biogenic synthesis (green synthesis) of palladium nanoparticles having possible application as nanocatalyst in environmental remediation [44].

Gum Karaya: It is from *Sterculia urens* tree and is traditionally tapped by scarring the bark in trunk or at the base of the trunk with an axe. Ethaphon, a plant growth hormone has been reported useful in stimulating gum production without causing any

harm to plant health [45]. It is a partially acylated polysaccharide made of galacturonic acid, beta-D-galactose, glucuronic acid, L-rhamnose [46-48]. Karaya gum is one of the least soluble gums but it rapidly absorbs water (up to 100 times of its weight) to swell and forms viscous colloidal solutions [47]. At 1% concentration in water it forms a colloidal solution while at 4% a thick gel/paste. Unlike other gums, karaya gum is soluble in 60% alcohol, but insoluble in other organic solvents [47].

Karaya gum has many pharmaceutical uses as an adhesive for dental fixtures and ostomy-equipment, and as a base for salicylic acid patches [49, 50], corrective for blood sugar and plasma lipid levels [51]. Its demulcent properties make it an important ingredient of lozenges used to relieve sore throat [48]. Due to its bulk forming ability it acts as laxative. It is also boasted for its aphrodisiac powers. Though antibacterial activity of Karaya gum is not reported, it has been reported to reduce bacterial adhesion by 98% when applied as protective coating to dentures [52].

Similar to gum tragacanth, and ghatti gum, karaya gum has potential for biogenic synthesis of silver

[53] and copper [54] nanoparticles which possess useful antimicrobial activity against *E. coli*, *S. aureus* and *Micrococcus luteus* strains and have several potential therapeutic and pharmaceutical applications.

Conclusion: Plant gums have been consumed all over the world since centuries for their nutritional, culinary and therapeutic properties. Except a few examples, gums are usually not important as herbal antimicrobial agents, but have several therapeutic (tonic, anti-diarrhoeic, laxative, immune-stimulant, strength providers), industrial (thickeners, refining agents) and pharmaceutical (excipient, adhesives, binders) uses. Recent use of gums in green synthesis of nanomaterials from silver, copper and palladium etc., has paved the way to their utility in synthesis of antimicrobial nanomaterials and casings.

Acknowledgement: Authors are thankful for the Director ICAR-IVRI & ICAR-IINRG for permitting for the work and financial support throughout.

REFERENCES

- Smolinske SC. *CRC Handbook of Food, Drug, and Cosmetic Excipients*; CRC Press: New Delhi, **1992**.
- Vivas N et al. Effect of gum arabic on wine astringency and colloidal stability. *Prog Agric Viteo* 2001; 118(8): 175-6 [in French]
- Siddiqui MZ et al. 2015. Physico-chemical properties and protein profiling of gum exudates of *Acacia nilotica* collected from different agro-climatic zones in India. *Res J Chem Environ* 2015; 19(4): 29-36.
- Renard D et al. *Acacia senegal* gum: Continuum of molecular species differing by their protein to sugar ratio, molecular weight, and charges. *Biomacromol* 2006; 7 (9): 2637-49.
- Elkhalifa KF. *Forest Botany*; Khartoum University Press: Khartoum, **1996**.
- Sebastian MK, Bhandari MM. Medicinal plant lore of Udaipur district, Rajasthan. *Bull Med Ethnobot Res* 1984; 5: 122-34.
- Siddiqui MZ et al. *Buchanania Lanzas*: a species of enormous potentials. *World J Pharm Sci* 2014; 2(4): 374-9.
- Rajvaidhya S et al. Extraction, isolation and chemical structure elucidation of daidzein from bark of *Acacia arabica* (Lam.) Willd of Bhopal, Madhya Pradesh, India. *Int J Pharm Sci Res* 2014; 5(5): 2014-21.
- Baravkar AA et al. Pharmaceutical and biological evaluation of formulated cream of methanolic extract of *Acacia nilotica* leaves. *Res J Pharm Technol* 2008; 1(4): 481-3.
- Siddiqui MB, Husain W. Traditional treatment of diarrhoea and dysentery through herbal drug in rural India. *Fitoterapia* 1991; 62: 325-29.
- Banso A. Phytochemical and antibacterial investigation of bark extracts of *Acacia nilotica*. *J. Med Plants Res* 2009; 3: 82-5.
- Sharma C et al. *In vitro* evaluation of anti-microbial spectrum of *Acacia nilotica* leaves and bark extracts against pathogens causing otitis infection. *J Inno Biol* 2014; 1(1): 51-6.
- Lawrence R et al. Antibacterial Activity of *Acacia arabica* (Bark) Extract against selected multi drug resistant pathogenic bacteria. *Int J Curr Microbiol App Sci* 2015; 1(S): 213-22
- Dev SNC et al. Antimicrobial activity and phytochemical analysis of *Acacia nilotica* (L.) Del. *Indian J Appl Pure Bio* 2014; 29(2): 331-2.
- Das N, Chatterjee P. Evaluation of antimicrobial potentiality of 50% aqueous ethanolic leaf extract of *Acacia nilotica* Willd. *Asian J Pharma Clin Res* 2014; 7(1S): <http://innovareacademics.in/journals/index.php/ajpcr/article/viewFile/822/556> (accessed 17 July 2015).
- Vijayasanthi M et al. Evaluation of the Antibacterial Potential of various solvent extracts of *Acacia nilotica* Linn. leaves. *Hygeia J D Med* 2012; 4(1): 91-6.
- Bnuyan IA et al. *In Vitro* antimicrobial activity of gum arabic (Al Mannaand Tayebat) prebiotics against infectious pathogens. *Int J Pharma Pharmaceu Res Human* 2015; 3(3): 77-85.
- Sravani P et al. *In-vitro* experimental studies on selected natural gums and resins for their antimicrobial activity. *Res J Pharmaceu Biol Chem Sci* 2014; 5(1): 154-72.
- Rakshit M, Ramalingam C. Gum acacia coating with garlic and cinnamon as an alternate, natural preservative for meat and fish. *African J Biotechnol* 2013; 12(4): 406-13
- Gaitonde RV. Antimicrobial activity of metallic salts of Arabic acid. *Indian Drugs Pharmaceu Indust* 1979; 14: 39-41.
- Ahmed I et al. Screening of some Indian medicinal plants for their antimicrobial properties. *J Ethnopharmacol* 1998; 62: 183-93.
- Niratker C, Sailaja D. Preliminary phytochemical screening and evaluation of antimicrobial activity of *Buchanania lanzan* (chironji) from Chhattisgarh. *World J Pharmaceu Res* 2014; 3(9): 514-22.

23. Manjunath JR, Mithun NM. Antibacterial and antifungal activity of the *Buchanania lanzan* leaves. *Ethnopharmacol* 2011; http://inventi.in/journal/art_reader/3/9389/rapid/ethnopharmacology/pi (accessed on 15 July 2015).
24. Gentry HS. Gum Tragacanth in Iran. *Economic Bot* 1957; 11: 40-63.
25. Anonymus. Antimicrobial activity of biopolymers. http://shodhganga.inflibnet.ac.in:8080/jspui/bitstream/10603/36798/10/10_chapter%205.pdf (accessed on 15 July 2015).
26. Ranjbar-Mohammadi M *et al.* Fabrication of novel nanofiber scaffolds from gum tragacanth/poly(vinyl alcohol) for wound dressing application: in vitro evaluation and antibacterial properties. *Mater Sci Eng C Mater Biol Appl* 2013; 33(8): 4935-43.
27. Ghayempour S *et al.* Tragacanth gum as a natural polymeric wall for producing antimicrobial nanocapsules loaded with plant extract. *Int J Biol Macromole* 2014; 81: 514-20
28. Riccardi BA, Fahrenbach MJ. Hypocholesterolemic activity of mucilaginous polysaccharides in white leghorn cockerels. *Fed Proc* 1965; 24: 263-5.
29. Ochuba GU, von Riesen VL. Fermentation of polysaccharides by klebsiellae and other facultative bacilli. *Appl Environ Microbiol* 1980; 39: 988-92.
30. Yadav JP, Panghal M. *Balanites aegyptiaca* (L.) Del. (Hingot): A review of its traditional uses, phytochemistry and pharmacological properties *Int J Green Pharmacy* 2010; 1: 140-6.
31. Doughari JH *et al.* Antibacterial effects of *Balanites aegyptiaca* L. Drel. and *Morianga oleifera* Lam. on *Salmonella typhi*. *Afr J Biotechnol* 2007; 6: 2212-5.
32. Parekh J, Chanda S. In vitro screening of antibacterial activity of aqueous and alcoholic extracts of various Indian plant species against selected pathogens from Enterobacteriaceae. *Afr J Microbiol Res* 2007; 1: 92-9.
33. Otieno JN *et al.* The effect of local minerals Kadsaro towards the antimicrobial activity of medicinal plants extract. Case of Lake Victoria Basen, Tarim Tanzania. *Afr J Tradit Complement Alt Med* 2007; 4: 1-6.
34. Runyoro DK *et al.* Screening of Tanzanian medicinal plants for anti-*Candida* activity. *BMC Complement Alt Med* 2006; 30: 6-11.
35. Shahid M *et al.* Antimicrobial Potential of *Balanites aegyptiaca* (L.) Del, *Stevia rebaudiana* (Bert.) Bertoni, *tylophora indica* (Burm.f.) Merrill, and *Cassia sophera* (Linn.). *The Open Conf Procced J* 2012; 3: 63-9.
36. Hines DA, Eckman K. Indigenous multipurpose trees of Tanzania: Uses and economic benefits for people. Forestry Department, Corporate Document Repository of FAO. Working paper: FO:Misc/93/9. 1993; 276. <http://www.fao.org/docrep/019/x5327e/x5327e.pdf> (accessed on 18 July 2015)
37. Vimal OP, Tyagi PD. *Prosopis juliflora*: chemistry and utilization. In: Patel VJ, ed. *The Role of Prosopis in Wasteland Development*. Javrajbhai Patel Agroforestry Center, Surendrabag, Gujarat, India, 1986: OVP1-OVP8.
38. Vedak S, Raut SV. Study on antibacterial compounds from methanolic extract of bark of *Prosopis juliflora* (Vilayati babbul). *Int J Pharmaceu Sci Bus Manag* 2014; 2(6): 1-14 .
39. Thakur R *et al.* Evaluation of antibacterial activity of *Prosopis juliflora* (sw.) Dc. Leaves. *Afr J Tradit Complement Alt Med* 2014; 11(3): 182-8.
40. Castellani O *et al.* Hydrocolloids with emulsifying capacity. Part 2. Adsorption properties at the n-hexadecane–water interface. *Food Hydrocolloids* 2010; 24(2–3): 121–30.
41. Deshmukha AS *et al.* Gum ghatti: A promising polysaccharide for pharmaceutical applications. *Carbohydrate Polymers* 2012; 87: 980–6
42. Kaith BS *et al.* Temperature, pH and electric stimulus responsive hydrogels from Gum ghatti and polyacrylamide—synthesis, characterization and swelling studies. *Der Chemica Sinica* 2010; 1(2): 44–54.
43. Kora AJ, Sashidhar RB. Antibacterial activity of biogenic silver nanoparticles synthesized with gum ghatti and gum olibanum: a comparative study. *J Antibiotics* 2015; 68: 88-97.
44. Kora AJ, Rastogi L. Green synthesis of palladium nanoparticles using gum ghatti (*Anogeissus latifolia*) and its application as an antioxidant and catalyst. *Arabian J Chem* 2015. <http://dx.doi.org/10.1016/j.arabjc.2015.06.024> (accessed on 10 July 2015).
45. Nair MNB. Gum tapping in *Sterculia urens* Roxb. (Sterculiaceae) using ethephon. <http://www.fao.org/docrep/ARTICLE/WFC/XII/0148-B4.HTM> (accessed on 10 July 2015).
46. Tyler VE. *Pharmacognosy*; Lea & Febiger: Philadelphia, PA, 1981.
47. Leung AY. *Encyclopedia Of Common Natural Ingredients Used in Food, Drugs, and Cosmetics*; Wiley and Sons: New York, 1980.
48. Morton JF. *Major Medicinal Plants*; CC Thomas: Springfield, IL, 1977.
49. Evans WC. *Trease and Evans' Pharmacognosy*; 13th ed; Bailliere Tindall: London, 1989.
50. Bart BJ *et al.* Salicylic acid in karaya gum patch as a treatment for verruca vulgaris. *J Am Acad Dermatol* 1989; 20: 74-6.
51. Behall KM. Effect of soluble fibers on plasma lipids, glucose tolerance and mineral balance. *Adv Exp Med Biol* 1990; 270: 7-16.
52. Wilson M, Harvey W. Prevention of bacterial adhesion to denture acrylic. *J Dent* 1989; 17: 166-70.
53. Venkatesham M *et al.* A Novel green synthesis of silver nanoparticles using gum karaya: characterization, antimicrobial and catalytic activity studies. *J Cluster Sci* 2014; 25(2): 409-22.
54. Vellora V *et al.* Green synthesis of copper oxide nanoparticles using gum karaya as a biotemplate and their antibacterial application. *Int J Nanomed* 2013; 8: 889–98.