



Wilt-Causing Fungal Pathogens of Banana

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ABSTRACT

This study sought to utilize indigenous soil micro-organisms to suppress wilt-causing fungal pathogens of the banana. Fungal pathogens were isolated from wilt-affected rhizospheric soil, and potential antagonistic bacterial strains were isolated from healthy rhizospheric soil in the same area from which fungal pathogens were isolated. The antifungal activity of isolated micro-organisms against fungal pathogens was studied both in vitro and in vivo against fungal pathogens. It was found that *Fusarium oxysporum* and *Alternaria sp.* were pathogenic, while *Penicillium sp.*, *Bacillus velezensis* and *Bacillus subtilis* were antagonistic. Moreover, it was seen that *B. velezensis*, *B. subtilis* and *Penicillium sp.* inhibited the growth of the two fungal pathogens in both in vitro and in vivo experiments. An antagonistic consortium isolated in this study has demonstrated remarkable potential for controlling fungal diseases caused by *Fusarium sp.* and *Alternaria sp.* Therefore, the use of indigenous microflora to improve disease suppression of banana plants against soil-borne pathogens is a preferable approach.

Keywords: wilt-causing; fungal pathogens; banana

INTRODUCTION

Fusarium wilt is one of the most common diseases in all soil types around the world which results in a heavy economic reduction in most of the global banana production. These diseases are known to be a major biological limiting factor for the development of the present banana industry. Historically, the infection spread to the xylem of banana plantations and eventually destroyed most of the banana plant. Amani and Avagyan (2014) reported that *F. oxysporum* was isolated from

banana root, pseudo stem and fruit (wallery and robusta), which caused necrosis, wilt and rot also stated that *F. oxysporum* is a post-harvest fruit rot pathogen. Apart from *Fusarium* wilt the occurrence of another species of the pathogenic fungi *Alternaria sp.* that causes leaf disease that has been reported in infected banana plantations. The co-occurrence of both pathogens in bananas and their control has yet to be reported. To date, various approaches have been applied to cope with wilt control, including a chemical method based on specific fungicides, physical method biological

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control (application of bacteria, fungi such as *Trichoderma*, *Actinobacteria* and *Bacillus*, etc.) and breeding/selection of disease-resistant banana. Among them, physical and biological methods become attractive as they have no environmental impacts varieties. The use of biological control agents to protect and promote plant growth through colonization and multiplication in both rhizosphere and plant system could be a promising approach to manage wilt of bananas. However, this approach still has many difficulties to overcome. Indigenous micro-organisms are a group of endogenous microbial consortia that inhabit the soil, which can be used for environmental and agricultural purposes. Indigenous micro-organisms can express microbial antagonism and inhibit the growth and development of fungal pathogens in various ways, including the production of specific antifungal compounds (antibiosis), competition for growth, (myco)-parasitism and predation. Our work focuses on the biocontrol of banana wilt diseases caused by *Fusarium* sp. and *Alternaria* sp. using strains of indigenous micro-organisms isolated from disease-suppressive soil. Phytopathogenic fungi and their receptive antagonistic microbes were isolated from banana cultivation lands. The antagonistic effects of these isolates on phytopathogens were evaluated through in vitro antagonistic assays. In vivo pot trials were also conducted to evaluate the efficacy of a consortium of bio-control agents against the wilt diseases.

MATERIALS AND METHODS

Soil samples were collected from banana orchards at Guangxi Academy of Agricultural Science, Nanning, China. naturally wilt-affected banana rhizospheric soil and rhizospheric soil of healthy banana plants. The incidence of disease in sampled plantations was about 60% of total plants.

Samples were collected via random sampling method from 150 to 200 mm depth rhizospheric soil. Soil samples were packed in sterile bags and kept on ice during transportation and stored at 4°C for further studies. Isolation and identification of micro-organisms under study. Isolation and identification of pathogenic fungi, Isolation and identification of *Bacillus* strains

Methodology

Isolation and identification of pathogenic fungi

Fungal isolation was performed using potatoes dextrose agar (PDA) media supplemented with Rose Bengal (0.02 g l⁻¹) and chloramphenicol (50mg l⁻¹) (Gil et al. 2009) to inhibit the bacterial growth. In detail, 1 g of rhizospheric soil around the wilt-affected banana plant was collected and transferred to a 250 ml conical flask containing 100 ml of sterile distilled water (Pitt and Hocking 1988;

Leslie and Summerell 2006; Woudenberg et al. 2013). After shaking the soil solution for 10 min at 120 rev min⁻¹, 1 ml of the supernatant was taken and diluted serially (107 times), after which 1 ml of each dilution was poured into the prepared PDA medium.

Isolation and Identification of *Bacillus* strains

Bacterial strains were isolated from rhizospheric soil around healthy banana plants using Luria–Bertani (LB) medium, supplemented with cyclohexamide (50 mg l⁻¹) to inhibit fungal growth, by serial dilution method like the method for serial dilution of fungal strains (refer to Isolation and identification of pathogenic fungi section) (Weisburg et al.1991). After 48 h incubation, bacterial isolates were enumerated and differentiated by colonial morphology (i.e., size, shape and colour; Thangavelu 2015). Individual colonies were purified by streaking on LB agar medium. For further analysis, bacterial isolates' glycerol stock cultures were maintained at -80°C.

Screening of potential antifungal properties of isolated antagonist strains

In order to screen the potential antifungal activity of isolated bacterial strains for their potential use as biocontrol agents (Zivković et al. (2010), three methods were followed, which are: 1) Dual-plate assay (competitiveness), Scanning electron microscopic studies, Effect of metabolite extracts on fungal growth inhibition (antibiosis), Fungal growth inhibition study.

In vivo experimental conditions

All pots were watered to saturation and placed in a greenhouse at Beijing University of Chemical Technology (39°58'02"N, 116°25'02"E), receiving 12 h sunlight each day with fluctuating day temperatures of 30–35°C and 20–24°C at night. The plantlets were watered once a day from the second day onwards, and no fertilizer was applied to the soil. After the 9th week of infection and treatment, wilt severity assessments were recorded. All plants' disease symptoms were evaluated according to a five-class variation from 0 to 4. 0-no yellowing of the leaves, the plant looks healthy; 1-light yellowing of the lower leaves; 2-yellowing of the lower leaves; 3-yellowing of most or all the leaves and 4-dead plant.

RESULTS

Total fungal count in the wilted rhizospheric soil was found to be 2.4×10^4 CFU per g. Of numerous colonies, three colonies, i.e., white cottony colony, white colony with green margin and thick colony which has dark green colour, were found to be dominating in the soil sample with distinct morphology. These colonies were selected and

subculture to test their pathogenicity in vivo. In vivo investigation of the pathogenicity of the identified fungal strains on banana plantlet and fruits was performed (as illustrated in (Fig. 1a-c)). In vivo experiments with single isolates of *B. subtilis*, *B. velezensis* or *Penicillium* sp. were shown to suppress wilt development without any statistical significance. It was observed that in the in vivo experiments, the combination of isolates *B. subtilis*, *B. velezensis* and *Penicillium* sp. reduced the disease severity to the greatest degree and controlled both phytopathogens up to 60–63%. The combined application of *B. subtilis* + *B. velezensis*

+ *Penicillium* sp. gave a significant reduction of disease severity among the tested combinations. This combination significantly reduced the disease severity and increased control. Control plants that were infected with both *F. oxysporum* and *Alternaria* sp. showed the characteristic wilt symptoms, and the disease severity index reached as high as 84–92%. It also showed that there was no significant difference between the tested isolates when used individually. In contrast, the combination of all isolates, disease severity was significantly lower than the untreated control plants.

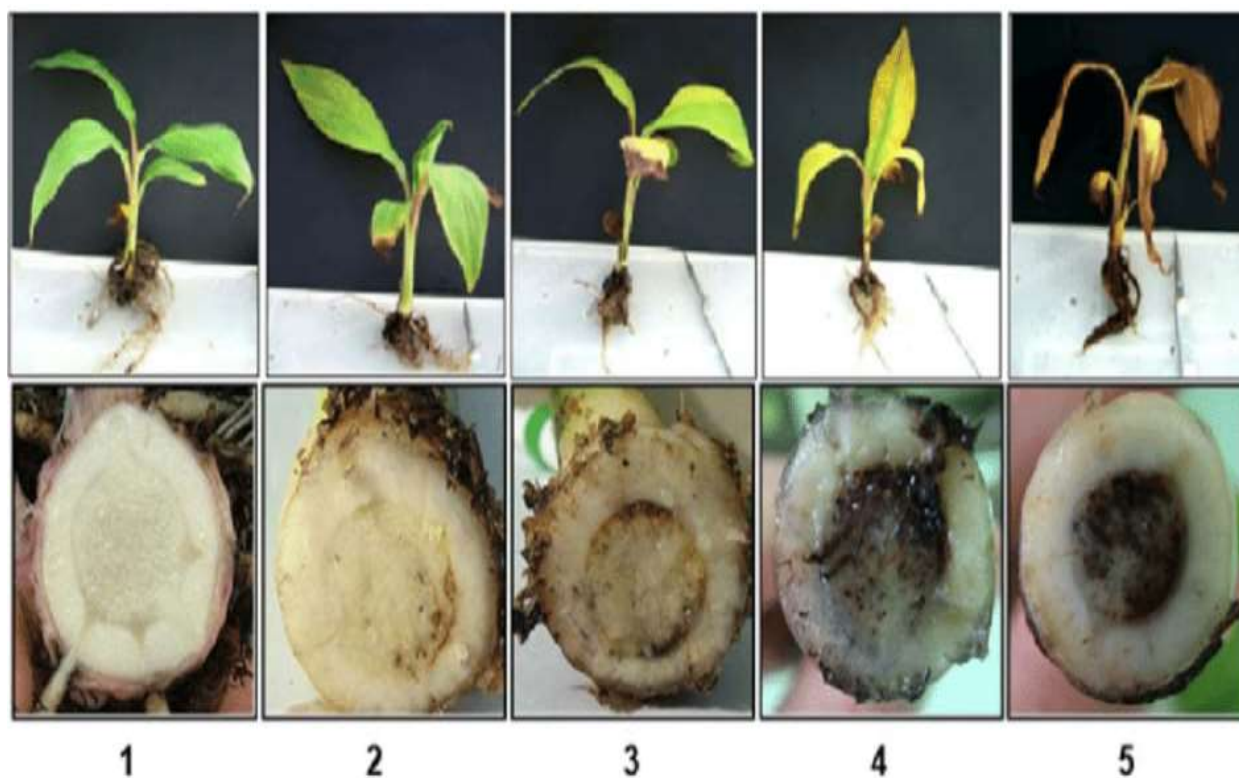


Figure 1: Scale for evaluation of *Fusarium* wilt of banana in greenhouse conditions based on external (upper panel) and internal symptoms (lower panel). Classes for external symptoms are: 1: No symptoms; 2: Initial yellowing mainly in the lower leaves; 3: Yellowing of all the lower leaves with some discoloration of younger leaves; 4: All leaves with intense yellowing; 5: Plant dead. Class for internal symptoms are: 1: No symptoms; 2: Initial rhizome discoloration; 3: Slight rhizome discoloration along the whole vascular system; 4: Rhizome with most of the internal tissues showing necrosis; 5: Rhizome totally necrotic. Source: http://www.scielo.org.mx/scielo.php?script=sci_arttext&pid=S0187-5779202000030043

DISCUSSION

To establish an effective control and management system, identifying the disease-causing agents is the preliminary necessity (Miller et al. 2009). In this study, two fungal isolates (*F. oxysporum* and *Alternaria* sp.) have been introduced for their pathogenicity with banana plantlets and fruits. Their macro- and micro-morphological structure were also consistent with the previous studies (Leslie and Summerell 2006; Woudenberg et al. 2013).

The observed disease symptoms were consistent with the description of symptoms shown by Jamil et al. (2019). The observed smaller yellowing surfaces as formed by *F.oxysporum* compared to *Alternaria* sp. could be explained as *F.oxysporum* predominantly penetrates roots, but not photosynthetic tissues (Stover 1962; Avagyan 2014). In addition, *Alternaria* sp. directly penetrates the host, through wounds or through stomata (Laemmlen 2002). Once the disease has been identified, bacterial antagonists were screened

against *Fusarium* sp. and *Alternaria* sp. For their biocontrol activity (Zivkovi'c *et al.* 2010). The present work made an attempt to isolate antagonist bacterial strains that could be used as biocontrol agents. The fungal isolate identified here as *Penicillium* sp. further showed the macro- and microscopic structure. Another interesting observation was that the metabolite extract of the antagonist bacterial strains could alter the growth of mycelium of the pathogenic fungi from 38% to 56% after 7 days of incubation at 28 °C. Growth of pathogens was inhibited in the presence of these antagonistic metabolites extract. The reduction in growth via metabolite extract of antagonist bacteria may be noted with the corresponding potential suppression of the fungal pathogen activity in the soil. It can also be assumed that the antagonists could reduce the invasion and subsequent development of pathogenic fungi, suggesting a high potential for biocontrol ability. Chitinase and β -1,3-glucanase are the key enzymes associated with the decomposition of the fungal cell wall, involved in myco-parasitism. These enzymatic activities have thus been previously used to evaluate the antifungal potential of various biological antifungal agents (Mendoza and Sikora 2009; Tan *et al.* 2015; Kohl *et al.* 2019). This work has also focused on the activities of chitinase and β -1,3-glucanase by growing the antagonist microorganisms under non-induced conditions, i.e. LB medium, without any

supplementation. The final disease control effect was between 24 and 63% in our *in vivo* study. These findings showed the ability of the isolated strains to survive in the host rhizospheric soil and the isolates' adaptability to the soil environment. Thus, the antagonist isolates depict the potential to control banana disease under field conditions. However, their co-existence as a consortium must be confirmed by an *in vitro* compatibility test. Further research on the biocontrol activity of these strains must be attempted with an enhancement of antifungal enzymes production from biocontrol agents and increasing resistance of biocontrol agents against environment constraints. In this study, two bacterial isolates and one fungal isolate showed a significant reduction in disease incidence against isolated phytopathogens. In particular, the mixture of all three isolates showed significance in disease suppression for both *F. oxysporum* and *Alternaria* sp. The use of indigenous microflora for the improvement of disease suppression of banana plants against soil-borne pathogens is a preferable approach, and further research is needed. These results support further investigation of the modes of action of the biocontrol agents at the molecular level, for example, via mutation studies focus on enhancement of antifungal enzymes production from biocontrol agents and increasing resistance of biocontrol agents against environment constraints.

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